Supplementary Materials:

CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells

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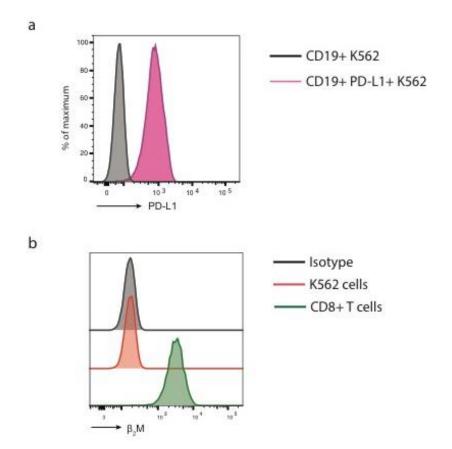
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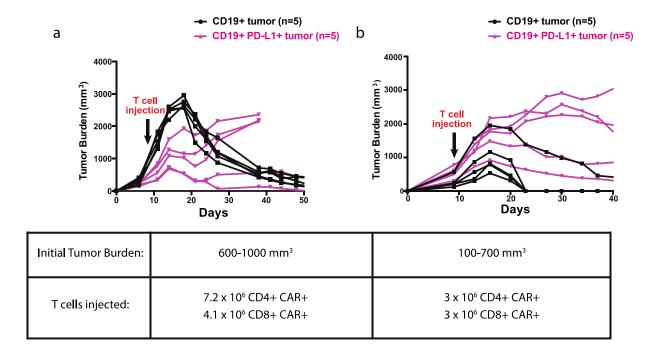
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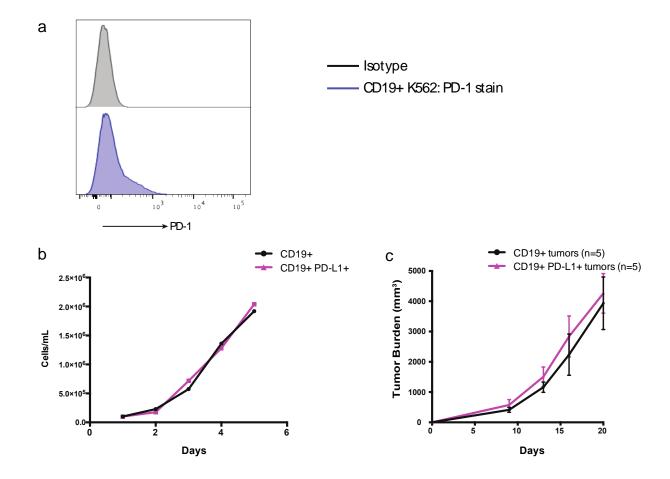
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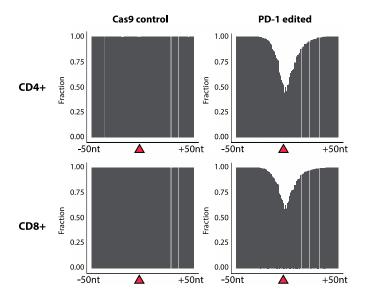
Supplementary Fig. 1. Characterization of CD19+ PD-L1+ K562 cell line. (a) Surface expression of human PD-L1 on transduced or parental CD19+ K562 cell lines. (b) Surface expression of beta-2-microglobulin (β_2 M) in CD19+ K562 cells and control CD8+ T cells as assayed by flow cytometry.



Supplementary Fig. 2. Impaired clearance of CD19+ PD-L1+ tumor xenografts across multiple initial tumor burdens and anti-CD19 CAR T cell doses. (a) NOD-scid-IL-2R $\gamma^{-/-}$ (NSG) mice were injected with 5 x 10⁶ CD19+ or CD19+ PD-L1+ K562 cells subcutaneously. Tumors were established at 600-1000 mm³ and mice were treated with 7.2 x 10⁶ CD4+ and 4.1 x 10⁶ CD8+ anti-CD19 CAR+ T cells. Tumor burden was measured longitudinally by caliper; curves show individual mice (n=5 per group). (b) NOD-scid-IL-2R $\gamma^{-/-}$ (NSG) mice were injected with 5 x 10⁶ CD19+ or CD19+ PD-L1+ K562 cells subcutaneously. Tumors were established at 100-700 mm³ and mice were treated with 3 x 10⁶ CD4+ and 3 x 10⁶ CD8+ anti-CD19 CAR+ T cells. Tumor burden was measured longitudinally by caliper; curves show individual mice (n=5 per group).

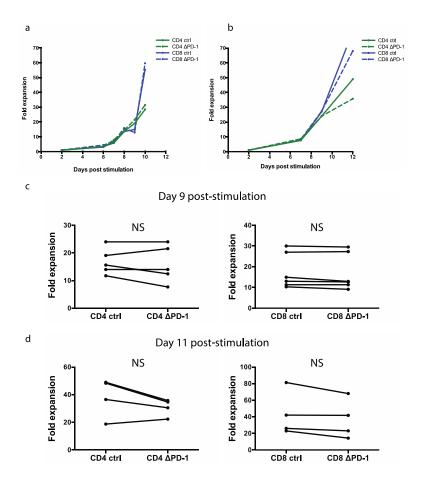


Supplementary Fig. 3. PD-1 expression and growth kinetics of CD19+ and CD19+ PD-L1+ cells lines. (a) Surface PD-1 expression in CD19+ K562 cultures as assayed by flow cytometry. (b) *In vitro* growth kinetics of CD19+ and CD19+ PD-L1+ cells. 1 x 10⁵ cells/mL were seeded on day 0 and cells were counted daily for 5 days. (c) *In vivo* growth kinetics of CD19+ and CD19+ PD-L1+ cells. NSG mice were injected subcutaneously with 5 x 10⁶ CD19+ or CD19+ PD-L1+ K562 cells and tumor burden measured longitudinally by caliper. Tumor burdens are mean ±S.D.

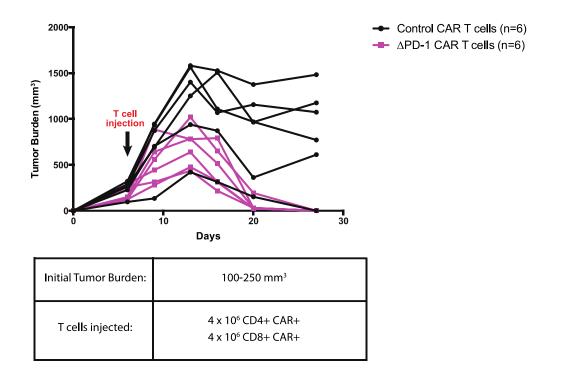


Supplementary Fig. 4. Deep sequencing of Pdcd1 locus in control and PD-1 edited cells.

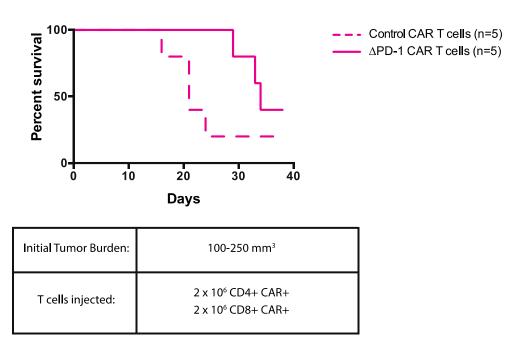
Deep sequencing of the PD-1 target site in Cas9 control or PD-1 edited cells. Fraction represents the proportion of reads mapping to the reference sequence for a given nucleotide in a 100 base window spanning the cut site (denoted by **\(\Delta \)**).



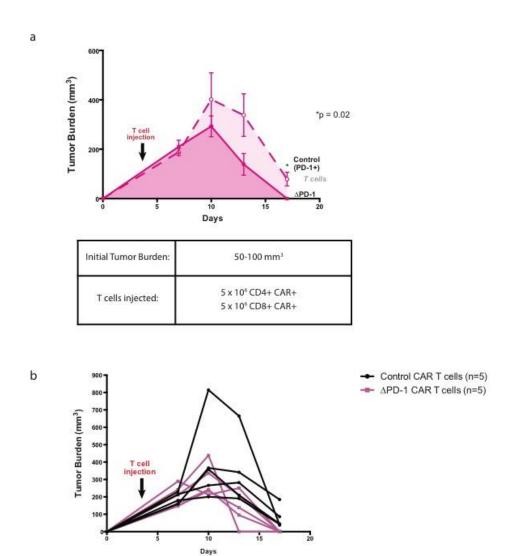
Supplementary Fig. 5. No impairment of CAR T cell expansion following *Pdcd1* editing via CRISPR. (a) Fold expansion of Cas9 control and PD-1 edited CD4+ or CD8+ T cells during representative experiment with no difference in expansion between treatment conditions. (b) Fold expansion of Cas9 control and PD-1 edited CD4+ or CD8+ T cells during representative experiment with reduced expansion of PD-1 edited cells following stimulation. (c) Fold expansion of Cas9 control and PD-1 edited CD4+ or CD8+ T cells 9 days after stimulation across multiple experiments. NS = not statistically significant; paired t-test. (d) Fold expansion of Cas9 control and PD-1 edited CD4+ or CD8+ T cells 11 days after stimulation across multiple experiments. NS = not statistically significant; paired t-test.



Supplementary Fig. 6. Improved clearance of CD19+ PD-L1+ tumor xenografts with PD-1 edited CAR T cells. Longitudinal tumor burdens for individual mice from experiment shown in Fig. 3d. NSG mice were injected with 5 x 10⁶ CD19+ PD-L1+ K562 cells subcutaneously. Mice with established tumors (100-250 mm³) were injected intravenously with 4 x 10⁶ CD4+ CAR+ and 4 x 10⁶ CD8+ CAR+ control T cells or PD-1 edited cells, and tumor burden measured longitudinally by caliper.



Supplementary Fig. 7. Kaplan-Meier curve of NSG mice with 100-250 mm³ initial tumors treated with 2 x 10⁶ CD4+ and 2 x 10⁶ CD8+ control or PD-1 edited anti-CD19 CAR+ T cells. NSG mice were injected with 5 x 10^6 CD19+ PD-L1+ K562 cells subcutaneously. Mice with established tumors (100-250 mm³) were injected intravenously with 2 x 10^6 CD4+ CAR+ and 2 x 10^6 CD8+ CAR+ control T cells or PD-1 edited cells.



Supplementary Fig. 8. Accelerated clearance of 50-100 mm³ tumors with PD-1 edited CAR T cells. (a) NSG mice were injected with 5 x 10⁶ CD19+ PD-L1+ K562 cells subcutaneously. Mice with 50-100 mm³ tumors were injected intravenously with 5 x 10⁶ CD4+ CAR+ and 5 x 10⁶ CD8+ CAR+ control T cells or PD-1 edited cells, and tumor burden measured longitudinally by caliper. Tumor burdens are mean ±SEM for each group (n=5 mice per group). 100% of animals receiving PD-1 edited CAR T cells cleared tumors by day 18, versus 0% in the control group (*p=0.02, Student's t-test). (b) Longitudinal tumor burdens for individual mice from experiment shown in (a).